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NEW EUDESMENOIC ACID METHYL ESTER FROM *ARTEMISIA SELENGENSIS*

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ABSTRACT

The aerial parts of *Artemisia selengensis* yielded a new eudesmenoic acid methyl ester together with a known eudesmanolide. The structures were elucidated by spectroscopic methods. The relative stereochemistry of the title compound was further supported by the result of a successful single-crystal X-ray diffraction analysis.

Key Words: *Artemisia selengensis*; Compositae; Sesquiterpene; Eudesmenoic acid methyl ester; Structural elucidation; X-ray diffraction analysis

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INTRODUCTION

Artemisia selengensis Turcz (family Compositae, tribe Anthemideae), known in China as "Hong-Chen-Ai," is a species of the well-known traditional Chinese medicine (TCM) "Liu-Ji-Nu." It grows wild in the southwestern part of China and is used locally for anti-inflammation, hemostasis, invigorating the blood circulation, and relieving dysmenorrhea (1). In a preliminary communication (2), we have reported two new guaianolides from the ethanolic extract of the aerial parts of *A. selengensis*. In addition to a known eudesmanolide douglanin **1**, we hereby describe the isolation and structure identification of a new eudesmene **2**, which was obtained during our further chemical investigation of this plant species. The structure of the latter compound was determined to be 1 α ,6 α -dihydroxyeudesma-3,11(13)-dien-12-carboxylic acid methyl ester by spectroscopic methods (IR, UV, EIMS, ^1H NMR, ^{13}C NMR, DEPT, and ^1H - ^1H COSY). The relative stereochemistry of **2** was further supported by the result of a successful single-crystal X-ray diffraction analysis.

EXPERIMENTAL

All 1D and 2D NMR spectra were measured on a Bruker AM-500 spectrometer, at room temperature, using TMS as an internal standard; chemical shifts are expressed in ppm. IR spectra were recorded on a Perkin-Elmer 683 spectrometer. EIMS (at 70 eV) were collected on a direct probe on an Analytical VG ZAB-2F. UV spectra were scanned on a UV-240 (Shimadzu) spectrophotometer. Optical rotations were determined on a Perkin-Elmer 241 polarimeter. For elemental analysis, we used a Carlo Erba-1106 apparatus. Melting points were taken on a Kofler hot-stage apparatus and are uncorrected. Silica gel (200–300 mesh) was used for column chromatography (CC), and for TLC we used precoated with silica gel (0.25 mesh) F₂₅₄. Spots were visualized using UV light (254 nm) and 5% H₂SO₄-EtOH.

The aerial parts of *A. selengensis* were collected from Peng County in the Si-Chuan province of P. R. China in November 1990. They were identified by Prof. Y. H. Chen of the Department of Botany in our Institute (IMM). A voucher specimen was deposited at the herbarium of the Institute of Chinese Herb in Peng County.

The dried and pulverized aerial parts (5 kg) of *A. selengensis* were extracted with petroleum ether (60–90°C) in a percolator, and the defatted material was exhaustively extracted with hot 95% EtOH (10 L). Ethanol extract was concentrated under reduced pressure to give a black residue (300 g), 120 g of which was suspended in 0.5 L of boiling H₂O and filtered. Then the aqueous filtrate was extracted with petroleum ether (60–90°C), EtOAc and *n*-BuOH, respectively. The



EtOAc extract (8 g) was chromatographed on a silica gel column (200–300 mesh, 250 g) eluted with CHCl_3 –MeOH gradient as the developing solvent. The eluent was monitored by TLC and combined to give 11 fractions. Compound **1** (5 mg) and compound **2** (12 mg) were obtained from fr. 7 (CHCl_3 /MeOH, 10:1) and purified by repeated silica gel CC and by preparative TLC.

Douglanin (1). Light yellow gum; IR (dry film) ν_{max} 3417 (OH), 2920, 2872, 1766 ($\text{C}=\text{O}$, α,β -unsaturated- γ -lactone), 1652 ($\text{C}=\text{C}$), 1456, 1375, 1249, 1137, 1045, 964 cm^{-1} ; UV (MeOH) λ_{max} ($\log \epsilon$): 208 (3.85) nm; EIMS m/z (rel. int.): 248 ($[\text{M}]^+$, 5), 230 ($[\text{M} - \text{H}_2\text{O}]^+$, 100), 215 ($[\text{M} - \text{H}_2\text{O} - \text{CH}_3]^+$, 53); ^1H NMR spectral data: see Table 1.

1 α ,6 α -Dihydroxyeudesma-3,11(13)-dien-12-carboxylic acid methyl ester (2). Colorless crystals (MeOH), m.p. 201–202°C; $[\alpha]_{\text{D}}^{16}$: +46° (*c* 0.04, MeOH); UV (MeOH) λ_{max} ($\log \epsilon$): 210 (3.91) nm; IR (KBr) ν_{max} : 3319 (OH), 2949, 2881, 1710 ($\text{C}=\text{O}$, α,β -unsaturated carboxylic acid ester), 1624 ($\text{C}=\text{C}$), 1436, 1326, 1149, 1043, 945 cm^{-1} ; EIMS m/z (rel. int.): 280 $[\text{M}]^+$ (not found), 262 ($[\text{M} - \text{H}_2\text{O}]^+$, 7), 244 ($[\text{M} - 2\text{H}_2\text{O}]^+$, 51), 230 (22), 215 (33), 197 (26), 185 (35), 169 (47), 107 (100), 91 (85), 79 (78), 67 (71), 55 (86), 53 (91), 43 (81); Elemental analysis (Anal. Calc. for $\text{C}_{16}\text{H}_{24}\text{O}_4$: C, 68.57; H, 8.57. Found: C, 68.45; H, 8.51%). ^1H NMR, ^{13}C NMR, and ^1H – ^1H COSY spectral data: see Table 1.

X-ray structure analysis of 2. $\text{C}_{16}\text{H}_{24}\text{O}_4$: A colorless crystal of **2** with dimensions $0.2 \times 0.3 \times 1.0 \text{ mm}^3$ was selected for X-ray analysis, orthorhombic, space group $\text{P2}_12_12_1$, unit cell dimensions: $a = 7.7870$ (1) Å, $b = 9.3530$ (2) Å, $c = 21.0570$ (1) Å, $V = 1533.6$ (4) Å³, $Z = 4$. Single-crystal X-ray diffraction data were collected by using a MAC Science DTP 2030K Image Plate with graphite monochromate, Moka radiation. There were 1555 reflections of which 1184 were observed; the position of 20 nonhydrogen atoms was obtained directly from E-map. The structure was solved with NOMCSDP software package; thereafter the positions of the nonhydrogen atoms were obtained and the kind of atoms was determined by using the least square calculation and the difference Fourier method in turn. Position of all hydrogen atoms was obtained by geometric calculation and difference Fourier method. The final reliable factors were $R_{\text{f}} = 0.061$, $R_{\text{w}} = 0.060$, $\text{GOF} = 5.837$.

RESULTS AND DISCUSSION

The minor component (**1**) gave rise to a clear molecular-ion peak at m/z 248, and the IR as well as the UV spectra displayed the presence of hydroxyl (IR: 3417 cm^{-1}) and α -methylene- γ -lactone (IR: 1766 cm^{-1} ; UV λ_{max} : 208 nm) groups. Its ^1H NMR spectrum (Table 1), reported here in detail for the first time, established the structure of **1** to be 1 α -hydroxyeudesma-3,11(13)-dien-12,6 α -olide. The structure of douglanin (**1**) was reported earlier (3).



Table 1. ^1H (500 MHz) NMR Data for **1**, **2** and ^{13}C - (125 MHz) NMR Data for **2**

Proton	δ , mult., J (Hz), int.			Observed ^1H - ^1H COSY, 2 ^b	Carbon	δ (DEPT), 2 ^b
	1 ^a	2 ^b				
H-1 β	3.34, brd, 4.0, 1H	3.32, brd, 4.7, 1H		H-2 β	C-1	72.07 (CH) ^c
H-2 α	2.05, brd, 19.1, 1H	2.05, brd, 18.8, 1H		H-2 β	C-2	34.85 (CH ₂) ^d
H-2 β	2.37, brd, 19.1, 1H	2.48, brd, ^e 18.8, 1H		H-1 β , H-2 α	C-3	121.17 (CH)
H-3	5.25, brs, 1H	5.29, brs, 1H		H-2 α , H-2 β , H-5 α	C-4	136.68 (C)
H-5 α	2.70, brd, 11.4, 1H	2.29, brd, 10.1, 1H		H-3, H-6 β	C-5	52.13 (CH)
H-6 β	4.00, dd, 10.8, 11.4, 1H	3.95, dd, 10.1, 10.5, 1H		H-5 α , H-7 α	C-6	74.43 (CH) ^c
H-7 α	2.51, ddt, 10.8, 5.1, 3.2, 3.0, 1H	2.48, m, ^e 1H		H-6 β , H-8 α , H-8 β	C-7	46.56 (CH)
H-8 α	1.68, m, 1H	1.87, m, ^f 1H		H-8 β , H-9 α , H-9 β	C-8	26.98 (CH ₂)
H-8 β	1.24, m, ^e 1H	1.63, ddt, 14.5, 11.1, 5.7, 1H		H-8 α , H-9 α , H-9 β	C-9	33.04 (CH ₂) ^d
H-9 α	1.91, m, 1H	1.90, m, ^f 1H		H-8 α , H-8 β , H-9 β	C-10	40.97 (C)
H-9 β	1.24, m, ^e 1H	1.16, ddd, 13.3, 5.7, 2.3, 1H		H-8 α , H-8 β , H-9 α	C-11	144.64 (C)
H-13	5.90, d, 3.2, 1H	6.27, d, 1.1, 1H		H-13'	C-12	169.22 (C)
H-13'	5.43, d, 3.0, 1H	5.75, brs, 1H		H-13	C-13	126.35 (CH ₂)
H-14	0.81, s, 3H	0.88, s, 3H			C-14	17.23 (CH ₃)
H-15	1.80, brs, 3H	1.88, brs, 3H			C-15	23.69 (CH ₃)
OMe		3.78, s, 3H			OMe	52.93 (CH ₃)

^aRecorded in acetone-d₆.

^bRecorded in CD₃OD.

^{c,d}Assignments in the same column with the same superscript may be interchangeable.

^{e,f}Proton signals were hidden or overlapped with the same superscript in the same column.

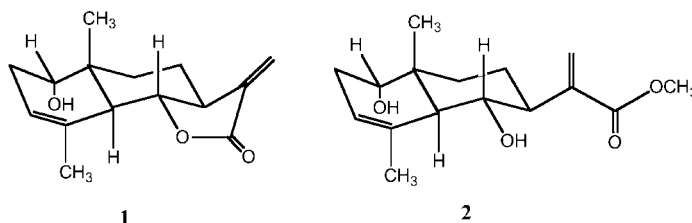


Figure 1. The structures of **1** and **2**.

The EI mass spectrum of compound **2** showed a clear $[M - H_2O]^+$ peak at m/z 262. This, together with its 1H and ^{13}C NMR (DEPT) spectral data (Table 1) and an elemental analysis, indicated the molecular formula to be $C_{16}H_{24}O_4$. The prominent mass spectral fragments and the 1H NMR spectrum data showed general features similar to those of $6\alpha,8\alpha$ -dihydroxyisocostic acid methyl ester obtained earlier by us from *A. mongolica* (4,5). This indicated that the structure of **2** is also an eudesmen-12-oic acid methyl ester skeleton: δ 0.88 (3H, brs, H-14), 1.88 (3H, brs, H-15), 6.27 (1H, d, $J = 1.1$ Hz, H-13), 5.75 (1H, brs, H-13') and 3.78 (3H, s, COOMe). A proton signal geminal to a hydroxyl group showed a broad doublet with a small coupling constant ($J = 4.7$ Hz) at δ 3.32. This clearly indicated that this proton was equatorial and β -orientated at C-1 on the basis of analogies with similar compounds such as douglanin (**1**) ($J_{1\beta,2\beta} = 4.0$ Hz) and $11\beta,13$ -dihydrodouglanin acetate ($J_{1\beta,2\beta} = 4.5$ Hz) (6).

For the presence of a double bond at C-3, ring A required a semichair conformation (Figs. 1 and 2). The downfield shift of H-2 β at δ 2.48 is in agreement with the proposed stereochemistry, and this proton is in the deshielding zone of the C-3 double bond. H-2 α proton signal at δ 2.05 is in the shielding zone of this double bond. Therefore, the dihedral angle between H-2 α and H-1 β is almost 90° ($J_{1\beta,2\alpha} = \sim 0$ Hz), so the proton signal of H-1 β showed a broad doublet coupling with H-2 β . The structure feature of H-1 β and its chemical shift assignment was further confirmed by 1H - 1H COSY spectrum, which revealed no

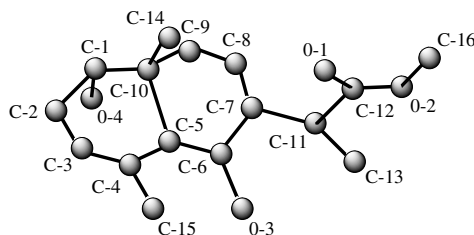


Figure 2. The X-ray structure of **2**.



cross peak between H-1 and other protons except for H-2 β (δ 2.48). We could assign unambiguously all the ^1H NMR signals by ^1H - ^1H COSY experiment (Table 1).

Furthermore, an evident explanation of this fact is that the molecule has a 1α -hydroxyl group that is responsible for the lowfield shifts of the H-5 α (δ 2.29) and H-9 α (δ 1.90) signals because 1,3-diaxial steric compression effect between 1α -OH and 5 α -H, as well as between 1α -OH and 9 α -H. The *trans*-decalin of the bicyclo (4,4,0) system followed from the biogenetic point of view. Therefore, the H-14 signal showed a singlet broadened by long range *W*-coupling between H-5 and H-14. The ^{13}C NMR and DEPT measurements were in full agreement with the proposed structure $1\alpha,6\alpha$ -dihydroxyeudesma-3,11(13)-dien-12-carboxylic acid methyl ester **2**. Finally, the relative stereochemistry of **2** was confirmed by X-ray diffraction for single crystals. Analysis results indicated that in the molecular skeleton of compound **2**, ring A (semichair) and ring B (chair) is linked by *trans*. Its relation configuration is displayed in Figure 1 and the X-ray structure in Figure 2. The absolute configuration remains unassigned. We went back to check the petroleum ether extract of the dried and pulverized material using TLC and NMR methods and found that both **1** and **2** are naturally occurring products.

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